



## How Do Foliar Application of Melatonin and L-Tryptophan Affect Lettuce Growth Parameters Under Salt Stress?

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| ARTICLE INFO   | ABSTRACT   |
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| <p><i>Research Article</i></p> <p>Received : 02/12/2019<br/>Accepted : 12/02/2020</p> <p><b>Keywords:</b><br/>Abiotic Stress<br/>Melatonin<br/>Seedling<br/>Tolerance<br/>L-Tryptophan</p> | <p>The aim of this study was to investigate the effects of exogenous Melatonin (100, 300 and 500 µM) and L-tryptophan (125, 250, 375 ppm) applications on some growth parameters of lettuce plants grown under salt stress. The study was carried out under semi-controlled greenhouse conditions in spring (March/April) season. The exogenous applications to lettuce plants were carried out two times as foliar spraying. Salt stress was generated by adding NaCl (0 mM, 100 mM, 200 mM) to irrigation water. The complete randomized design was used with three replications in this experiment. At the end of the study, it was found that the highest doses of exogenous applications had the highest effect on the parameters of the number of leaves, salinity necrosis, fresh leaf weight, fresh root weight, and total surface area of lettuce plants under 200 mM salinity condition. When the effects of the subtract on these values were compared, the effect of melatonin was found to be more pronounced. Leaf width, leaf length, and leaf surface temperature values were not affected by the external application. These values only changed depending on salt concentration. As a result of the study, it was concluded that the application of 500 µM melatonin significantly increased salt tolerance in lettuce plants. However, in order to reach a more general conclusion, the dose ranges and genotype/variety numbers should be increased.</p> |

Türk Tarım – Gıda Bilim ve Teknoloji Dergisi, 8(4): 960-964, 2020

## Yapraktan Uygulanan Melatonin ve L-Triptofan, Tuz Stresi Altındaki Marulun Büyüme Parametrelerini Nasıl Etkiler?

| MAKALE BİLGİSİ   | ÖZ   |
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| <p><i>Araştırma Makalesi</i></p> <p>Geliş : 02/12/2019<br/>Kabul : 12/02/2020</p> <p><b>Anahtar Kelimeler:</b><br/>Abiyotik stress<br/>Fide<br/>Melatonin<br/>Tolerans<br/>L-Triptofan</p> | <p>Bu çalışmanın amacı, dışsal Melatonin (100, 300 ve 500 µM) ve L-tryptophan (125, 250, 375 ppm) uygulamalarının, tuz stresi altında yetiştirilen marul bitkilerinin bazı büyüme parametreleri üzerine etkilerini incelemektir. Çalışma ilkbahar sezonunda yarı kontrollü sera koşullarında yürütülmüştür. Bitkilere dışsal uygulamalar yaprakтан spreyleme şeklinde iki kez yapılmıştır. Tuz stresi ise sulama suyuna ilave edilen NaCl (0 mM, 100 mM, 200 mM) ile oluşturulmuştur. Çalışma tesadüf parselleri deneme desenine göre üç tekerrürlü olarak yürütülmüştür. Çalışma sonunda, 200 mM tuz stresi koşulunda yetiştirilen bitkilerde yaprak sayısı, tuz nekrozu, yaprak yaş ağırlığı, kök yaş ağırlığı ve toplam yaprak yüzey alanı parametreleri üzerine en yüksek dozdaki dışsal uygulamaların, en yüksek etkiye sahip oldukları anlaşılmıştır. Bu değerler üzerine dışsal uygulamaların etkisi karşılaştırıldığında, melatonin etkisinin daha belirgin olduğu anlaşılmıştır. Yaprak genişliği, yaprak uzunluğu ve yaprak yüzey sıcaklığı değerlerine dışsal uygulamaların herhangi bir etkisi olmamıştır. Bu değerler sadece tuz konsantrasyonuna bağlı olarak değişmiştir. Çalışma sonucunda, 500 µM melatonin uygulamasının marul bitkisinde tuz toleransını önemli ölçüde artırdığı sonucuna varılmıştır. Ancak daha genel bir yargıya varmak için doz aralıklarının ve genotip/çesit sayısının artırılması gerektiği anlaşılmıştır.</p> |

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## Introduction

Lettuce (*Lactuca sativa* L.) is a one-year cool climate vegetable of the genus *Lactuca* of the *Compositae* (*Asteraceae*) family. Lettuce is among the most leaf consumed vegetables in the world. Lettuce has been cultivated for approximately 2500 years. Lettuce, which has been cultivated and fondly consumed for many years, can be found in markets throughout the year (Vural et al., 2000). According to the most recent data of the United Nations Food and Agriculture Organization (FAO), worldwide lettuce (and chicory) production was approximately 26.866.557 tonnes in 2017 from 1.227.358 hectares. According to the same data, Turkey produces 490.423 tonnes of lettuce (and chicory) annually, and it ranks as the 8th biggest producer (Anonymous, 2019).

Different ecological conditions of Turkey allow the cultivation of many types of vegetables. As a result of these advances, Turkey is the fourth-largest vegetable producer country after the United States, China, and India. Vegetable production has an important place in Turkey's economy. Also, Turkey is among the countries self-sufficient in terms of vegetable production (Yanmaz et al., 2015). However, this production potential is under various risks. Especially in greenhouse cultivation, soils have been faced with the threat of pollution and fatigue because of intensive plant production, monoculture applications, intensive use of chemicals, and the absence of natural rainfall. Stress factors in plants are grouped as biotic (viruses, bacteria, fungi, insects, herbivores, rodents, parasitic plants) and abiotic (salt, water, temperature, light, gases, minerals, radiation, etc.) stress factors (Ashraf and Foolad, 2007; Uyanık et al., 2014). The salinity which is one of the abiotic stresses is one of the most important reasons for the decrease in the yield of agricultural land (Tuteja, 2007). Salinity affects about 227 million hectares of irrigated agricultural land (Tuteja, 2007; Çulha and Çakırlar, 2012). Turkey is faced with 1.5 million hectares of land salinity. 60% of these lands are classified as saline, 19.6% as medium saline and 0.4% as alkaline (Kendirli et al., 2005). Salinity stress is an important issue for agricultural production and there are many different strategies to eliminate salinity stress (Ekbic et al., 2017). Exogenous application of plant growth regulators (PGRs) has proven quite successful in alleviating different types of abiotic stresses in plants (Ahmad et al., 2019).

Melatonin (Mel) was first obtained from the cattle brain gland (epiphysis) in 1958 (Lerner et al., 1958). Since the pineal tissue is found only in the nervous system of vertebrates and melatonin functions hormonal structure, this compound was initially thought to be a neural hormone (neurohormone) and synthesized only in the pineal tissue (Gundy et al., 1976). Melatonin was found in plants for the first time in 1995 by two different research groups as a result of independent research (Dubbels et al., 1995; Hattori et al., 1995). After the presence of plants, a growing number of researches have been conducted in different fruits, cereals, vegetables, seeds, medicinal and aromatic plants and ornamental and wild plant species (Paredes et al., 2009). Melatonin synthesis starts from tryptophan in all living organisms including plants, algae, and bacteria. Tryptophan is a compound from which the synthesis begins not for melatonin, but also serotonin and indole-3

acetic acid. The most widely accepted pathway for the synthesis of melatonin from tryptophan is Tryptophan-Tryptamine-Serotonin-Melatonin (Murch and Saxena, 2002).

L-tryptophan (3-indolylalanine) (L-tr), discovered by the British chemist Frederick Gowland Hopkins in 1901. It is an essential amino acid not only for plants but also for animals, humans and certain bacteria (Frankenberger and Arshad, 1991). It is a biologically active precursor molecule of auxin and therefore increases the level of auxin in plant tissues when exogenously applied. Although the positive effects of L-tr application on the germination and growth performance of various plants have been reported by several researchers (Abbas et al., 2013; Antony et al., 2017), there are no studies demonstrating their effects on lettuce.

While several studies have been published investigating the effects of different plant growth regulators and seed pretreatments on lettuce grown under abiotic stress, no studies have been found to investigate the role of L-tr or Mel applications together on growth parameters under salinity stress conditions. Therefore, the main objective of this study was to evaluate the effect of different L-tr and Mel concentrations on several parameters of lettuce plants under salinity conditions.

## Material and Methods

The study was carried out at the Erciyes University, Faculty of Agriculture, Kayseri, Turkey period of March-May, 2019. The experiment was conducted in semi-controlled greenhouse conditions with 3 replications by a completely randomized design.

All cultivation continued for about two months, during which the minimum temperature was 10°C and the maximum temperature was 18°C. Lettuce (*Lactuca sativa* L. cv. Grise Maraichère') was used as plant material. Seeds of lettuce were sown in viols which filled with perlite/peat (1:1) mixture. Twenty days old seedlings were then transplanted into two-liter pots. These pots were filled with 1.8 liters of perlite/peat/ soil mixture (1:1:1). Salt stress treatments were carried out as described by Ahmed et al. (2019) with little modifications. This experiment consisted of three treatments of NaCl (0 mM, 100 mM, 200 mM). The application of saline irrigation began one week after the seedlings were transplanted. The pots of each group were then divided into seven subgroups. Different levels of Mel (100, 300 and 500 µM) and L-tr (125, 250, 375 ppm) were applied to sub-groups. These agents were applied two times to 34 and 41 days old plants, as a foliar spray. All plants were sprayed with 30 ml of solution. Irrigation continued for five days' periods until the end of the trial (60 days). All agronomic applications were made as required. Data on twelve traits were recorded on an individual basis from three plants randomly chosen in each treatment. A scale of 0-5 was used in order to express the visible signs of damage caused by salt stress on lettuce plants. At the end of the experiment, standard discs were taken from middle-aged and young leaves of lettuce plants. The fresh weights of these discs and the fresh weights of all leaves were recorded. The total leaf area of the plant

was calculated using the obtained data. To determine the fresh and dry weights (g /plant) of the leaves and roots, each plant was weighed on a sensitive balance, then re-weighed after drying in a 65°C oven for 48 hours.

Leaf temperatures were measured from the three points of the widest leaf with an infrared thermometer. Statistical analysis was conducted using the JMP software. The data from the experiment were subjected to a general analysis of variance (ANOVA).

## Results and Discussion

The combined analysis of variance indicated that the effect of salinity statically significant for all measured characters. Furthermore, the interaction between experimental factors was significant for eight traits: number of leaves (LN), leaf color (LC), salinity necrosis (SN), total leaf fresh weight ( $\Sigma$ FLW), total fresh root weight ( $\Sigma$ FRW), total leaf surface area ( $\Sigma$ LSA), total dry leaf weight ( $\Sigma$ DLF), and total dry root weight ( $\Sigma$ DRW) (Table 1 and 2). Generally, all morphological parameters were reduced by increased salinity level except for salinity necrosis and leaf surface temperature.

The number of leaves ranged from 8.50 to 18.50 per plant. Although the number of leaves decreased with respect to the degree of the salinity, the doses of Mel and L-tr affected the severity of this decreasing. In lettuce plants not irrigated with salt water, the highest number of leaves was obtained in 250 ppm L-Tryptophan application.

In the highest salt stress condition (200 mM), application of 500  $\mu$ M Mel and 375 ppm L-tr resulted in a significant increase in the number of leaves compared to the control. Xu and Mou (2015) reported a decrease in fresh mass and dry mass values of 178 lettuce cultivars depend on salt stress. The present study showed a gradual reduction in plant weight and average leaf number with increasing salt concentrations consistent with the study as previously reported (Al-Maskri et al., 2010, Garrido et al., 2014)

Leaf color was determined visually. For this purpose, the color darkness was scored linearly from 1 to 5. While the color was darker in the plants without salt stress (4.07), the color was lightened due to increased salt concentration (3.64). Necrosis in leaves due to salinity stress was also visually scored. During this process, stress-free plants were used as control. In terms of these traits, 500  $\mu$ M Melatonin and 375 ppm L-Tryptophan applications showed very positive results under 200 mM salinity conditions. When the decreases in the averages due to increasing salt dose were examined, the most dramatic decrease was observed in  $\Sigma$ FLW and  $\Sigma$ LSA values (84%). In plants without salt stress, only the lowest doses of exogenous treatments caused an increase in  $\Sigma$ FLW values. No significant change was observed in 100 mM salt stress depending on applications. However, at the 100 mM stress conditions, 500  $\mu$ M Melatonin and 375 ppm L-Tryptophan applications significantly increased weight of fresh leaves values compared to control in the same group.

Table 1. Some growth parameters of lettuce plants at different salinity conditions

| Salinity | Treatment                 | LN                   | LC                | SN                | $\Sigma$ FLW         | $\Sigma$ FRW        | $\Sigma$ LSA          |
|----------|---------------------------|----------------------|-------------------|-------------------|----------------------|---------------------|-----------------------|
| 0        | 0                         | 14.50 <sup>A-E</sup> | 5.0 <sup>A</sup>  | 0 <sup>D</sup>    | 101.00 <sup>B</sup>  | 7.45 <sup>CD</sup>  | 2077.29 <sup>B</sup>  |
|          | 100 $\mu$ M Mel           | 16.50 <sup>AB</sup>  | 4.0 <sup>B</sup>  | 0 <sup>D</sup>    | 131.22 <sup>A</sup>  | 8.60 <sup>BC</sup>  | 2698.83 <sup>A</sup>  |
|          | 300 $\mu$ M Mel           | 14.50 <sup>A-E</sup> | 4.0 <sup>B</sup>  | 0 <sup>D</sup>    | 106.65 <sup>B</sup>  | 9.38 <sup>B</sup>   | 2193.39 <sup>B</sup>  |
|          | 500 $\mu$ M Mel           | 13.00 <sup>B-F</sup> | 4.0 <sup>B</sup>  | 0 <sup>D</sup>    | 107.13 <sup>B</sup>  | 5.91 <sup>DE</sup>  | 2203.36 <sup>B</sup>  |
|          | 125 ppm L-tr              | 13.50 <sup>B-E</sup> | 3.5 <sup>BC</sup> | 0 <sup>D</sup>    | 135.04 <sup>A</sup>  | 11.83 <sup>A</sup>  | 2777.29 <sup>A</sup>  |
|          | 250 ppm L-tr              | 18.50 <sup>A</sup>   | 4.0 <sup>B</sup>  | 0 <sup>D</sup>    | 92.65 <sup>B</sup>   | 5.66 <sup>DE</sup>  | 1905.55 <sup>B</sup>  |
|          | 375 ppm L-tr              | 13.00 <sup>B-F</sup> | 4.0 <sup>B</sup>  | 0 <sup>D</sup>    | 90.22 <sup>B</sup>   | 4.61 <sup>E-G</sup> | 1855.57 <sup>B</sup>  |
|          | Mean                      | 14.79                | 4.07              | 0.00              | 109.13               | 7.63                | 2244.47               |
| 200 mM   | 0                         | 16.00 <sup>A-C</sup> | 4.0 <sup>B</sup>  | 0 <sup>D</sup>    | 60.09 <sup>C</sup>   | 3.73 <sup>F-I</sup> | 1235.88 <sup>C</sup>  |
|          | 100 $\mu$ M Mel           | 15.50 <sup>A-D</sup> | 4.0 <sup>B</sup>  | 0 <sup>D</sup>    | 61.45 <sup>C</sup>   | 5.26 <sup>EF</sup>  | 1263.85 <sup>C</sup>  |
|          | 300 $\mu$ M Mel           | 14.50 <sup>A-E</sup> | 4.0 <sup>B</sup>  | 0.5 <sup>CD</sup> | 51.33 <sup>CD</sup>  | 3.14 <sup>G-J</sup> | 1055.61 <sup>CD</sup> |
|          | 500 $\mu$ M Mel           | 14.00 <sup>B-E</sup> | 4.0 <sup>B</sup>  | 0.5 <sup>CD</sup> | 64.80 <sup>C</sup>   | 4.46 <sup>E-G</sup> | 1332.75 <sup>C</sup>  |
|          | 125 ppm L-tr              | 13.50 <sup>B-E</sup> | 4.0 <sup>B</sup>  | 0.7 <sup>D</sup>  | 58.81 <sup>C</sup>   | 4.56 <sup>E-G</sup> | 1209.56 <sup>C</sup>  |
|          | 250 ppm L-tr              | 12.00 <sup>C-G</sup> | 4.0 <sup>B</sup>  | 0.5 <sup>CD</sup> | 39.28 <sup>DE</sup>  | 2.94 <sup>G-J</sup> | 807.78 <sup>DE</sup>  |
|          | 375 ppm L-tr              | 11.50 <sup>D-H</sup> | 3.5 <sup>BC</sup> | 1.5 <sup>BC</sup> | 67.98 <sup>C</sup>   | 4.11 <sup>E-H</sup> | 1398.16 <sup>C</sup>  |
|          | Mean                      | 13.86                | 3.93              | 0.53              | 57.68                | 4.03                | 1186.23               |
| 400 mM   | 0                         | 8.50 <sup>GH</sup>   | 3.5 <sup>BC</sup> | 3.5 <sup>A</sup>  | 6.85 <sup>H</sup>    | 2.42 <sup>H-J</sup> | 140.79 <sup>H</sup>   |
|          | 100 $\mu$ M Mel           | 7.50 <sup>H</sup>    | 3.0 <sup>C</sup>  | 3.5 <sup>A</sup>  | 9.76 <sup>GH</sup>   | 0.52 <sup>K</sup>   | 200.63 <sup>GH</sup>  |
|          | 300 $\mu$ M Mel           | 10.50 <sup>E-H</sup> | 3.5 <sup>BC</sup> | 3.5 <sup>A</sup>  | 18.50 <sup>F-H</sup> | 1.88 <sup>L-K</sup> | 380.39 <sup>F-H</sup> |
|          | 500 $\mu$ M Mel           | 15.00 <sup>A-D</sup> | 4.0 <sup>B</sup>  | 1 <sup>CD</sup>   | 35.38 <sup>D-F</sup> | 4.35 <sup>E-G</sup> | 727.57 <sup>D-F</sup> |
|          | 125 ppm L-tr              | 9.00 <sup>F-H</sup>  | 4.0 <sup>B</sup>  | 1.5 <sup>BC</sup> | 13.66 <sup>GH</sup>  | 1.72 <sup>JK</sup>  | 280.85 <sup>GH</sup>  |
|          | 250 ppm L-tr              | 10.50 <sup>E-H</sup> | 3.5 <sup>BC</sup> | 2.5 <sup>B</sup>  | 14.35 <sup>GH</sup>  | 1.63 <sup>JK</sup>  | 295.14 <sup>GH</sup>  |
|          | 375 ppm L-tr              | 14.00 <sup>B-E</sup> | 4.0 <sup>B</sup>  | 0.5 <sup>CD</sup> | 26.14 <sup>E-G</sup> | 2.84 <sup>G-J</sup> | 537.63 <sup>E-G</sup> |
|          | Mean                      | 10.71                | 3.64              | 2.29              | 17.81                | 2.19                | 366.14                |
| F ratios | Salinity                  | 14.47*               | 5.60*             | 47.71*            | 428.86*              | 131.57*             | 428.86*               |
|          | Hormone                   | 0.55                 | 1.27              | 1.61              | 5.15*                | 5.17*               | 5.15*                 |
|          | Salinity $\times$ Hormone | 3.10*                | 2.57*             | 3.58*             | 4.77*                | 7.71*               | 4.77*                 |

Means in a column followed by the different letters are significantly different at the various levels as determined by LSD test. CV: coefficient of variation, LSD: least significance difference. P < 0.01. Mel: Melatonin, L-tr: L-Tryptophan, LN: Number of leaves, LC: Leaf color, SN: Salinity necrosis,  $\Sigma$ FLW: Total fresh leaves weight,  $\Sigma$ FRW: Total root fresh weight;  $\Sigma$ LSA: Total surface area.

Table 2. Some growth parameters of lettuce plants at different salinity conditions

| Salinity | Treatment          | LN                   | LC                   | SN                 | ∑FLW               | ∑FRW              | ∑LSA                |
|----------|--------------------|----------------------|----------------------|--------------------|--------------------|-------------------|---------------------|
| 0        | 0                  | 98.11 <sup>AB</sup>  | 8.49 <sup>C-E</sup>  | 14.75              | 20.00              | 7.50              | 143.97              |
|          | 100 µM Mel         | 124.52 <sup>A</sup>  | 9.77 <sup>A-D</sup>  | 14.40              | 20.50              | 7.50              | 165.36              |
|          | 300 µM Mel         | 108.35 <sup>A</sup>  | 7.90 <sup>D-F</sup>  | 14.40              | 21.50              | 10.00             | 169.49              |
|          | 500 µM Mel         | 101.40 <sup>A</sup>  | 7.35 <sup>D-F</sup>  | 14.50              | 20.75              | 9.50              | 208.30              |
|          | 125 ppm L-tr       | 108.70 <sup>A</sup>  | 12.08 <sup>A</sup>   | 14.00              | 19.00              | 6.50              | 143.61              |
|          | 250 ppm L-tr       | 107.72 <sup>A</sup>  | 9.72 <sup>A-D</sup>  | 14.45              | 20.50              | 7.50              | 151.41              |
|          | 375 ppm L-tr       | 108.40 <sup>A</sup>  | 10.90 <sup>A-C</sup> | 14.65              | 18.00              | 8.00              | 103.16              |
|          | Mean               | 108.17               | 9.46                 | 14.45 <sup>B</sup> | 20.04 <sup>A</sup> | 8.07 <sup>A</sup> | 155.04 <sup>A</sup> |
| 200 mM   | 0                  | 111.63 <sup>A</sup>  | 7.46 <sup>D-F</sup>  | 14.65              | 15.00              | 5.25              | 78.48               |
|          | 100 µM Mel         | 105.04 <sup>A</sup>  | 11.48 <sup>AB</sup>  | 14.55              | 17.50              | 6.25              | 82.12               |
|          | 300 µM Mel         | 44.25 <sup>CD</sup>  | 4.61 <sup>G-I</sup>  | 14.75              | 15.50              | 5.50              | 96.15               |
|          | 500 µM Mel         | 116.04 <sup>A</sup>  | 9.79 <sup>A-D</sup>  | 14.45              | 15.50              | 5.50              | 90.22               |
|          | 125 ppm L-tr       | 114.19 <sup>A</sup>  | 8.99 <sup>B-D</sup>  | 14.85              | 11.50              | 4.50              | 125.30              |
|          | 250 ppm L-tr       | 64.75 <sup>BC</sup>  | 6.12 <sup>E-G</sup>  | 14.30              | 16.25              | 6.50              | 72.49               |
|          | 375 ppm L-tr       | 23.52 <sup>DE</sup>  | 4.69 <sup>G-I</sup>  | 15.00              | 14.00              | 5.50              | 67.89               |
|          | Mean               | 82.77                | 7.59                 | 14.65 <sup>B</sup> | 15.04 <sup>B</sup> | 5.57 <sup>B</sup> | 87.52 <sup>B</sup>  |
| 400 mM   | 0                  | 7.78 <sup>E</sup>    | 4.04 <sup>G-I</sup>  | 15.55              | 8.75               | 2.50              | 16.75               |
|          | 100 µM Mel         | 10.13 <sup>E</sup>   | 2.36 <sup>I</sup>    | 14.95              | 9.00               | 2.75              | 27.62               |
|          | 300 µM Mel         | 22.69 <sup>DE</sup>  | 11.57 <sup>A</sup>   | 14.70              | 13.50              | 5.00              | 48.53               |
|          | 500 µM Mel         | 35.33 <sup>C-E</sup> | 6.01 <sup>E-G</sup>  | 14.70              | 7.25               | 2.25              | 38.24               |
|          | 125 ppm L-tr       | 26.84 <sup>DE</sup>  | 4.06 <sup>G-I</sup>  | 14.95              | 11.50              | 4.75              | 38.36               |
|          | 250 ppm L-tr       | 21.81 <sup>DE</sup>  | 3.35 <sup>HI</sup>   | 15.25              | 8.50               | 3.25              | 41.86               |
|          | 375 ppm L-tr       | 23.98 <sup>DE</sup>  | 5.76 <sup>F-H</sup>  | 15.35              | 9.50               | 3.50              | 27.80               |
|          | Mean               | 21.22                | 5.31                 | 15.06 <sup>A</sup> | 9.71 <sup>C</sup>  | 3.43 <sup>C</sup> | 34.17 <sup>C</sup>  |
| F ratios | Salinity           | 108.11*              | 40.58*               | 10.29*             | 68.71*             | 50.00*            | 123.44*             |
|          | Hormone            | 3.73*                | 2.20*                | 1.59               | 1.21               | 1.27              | 3.61                |
|          | Salinity × Hormone | 4.38*                | 11.67*               | 0.86               | 1.13               | 1.44              | 1.81                |

Means in a column followed by the different letters are significantly different at the various levels as determined by LSD test. CV: coefficient of variation, LSD: least significance difference. P < 0.01. Mel: Melatonin, L-tr: L-Tryptophan, ∑DLW: Total leaves dry weight, ∑DRW: Total root dry weight, LST: Leaves surface temperatures, LL: length of largest leaf, LW: Width of largest leaf, MLSA: Mean leaf surface area

In the salt-free group, total leaves surface area (TLSA) values were slightly increased with the lowest doses of exogenous applications, but no difference was observed in increasing doses. The results obtained at the highest salt dose (200 mM) in the TLSA examination showed parallels with other traits, and the highest external dose applications (500 µM Melatonin and 375 ppm L-Tryptophan) gave the best results. The fresh weights of the roots were not compatible with the dose increases of external applications in the salt-free medium. However, in the presence of severe salt stress, these values increased in parallel with the doses of exogenous applications.

Seed germination, fresh and dry weight of shoot, and root weight of lettuce have been affected both by ionic and osmotic effects due to salinity (Barassi et al., 2006). In a study carried out by Ahmed et al. (2019), depending on increasing salt doses, shoot length, root length, total plant weight, and leaf number were significantly decreased. The results obtained in our study are in parallel with these findings. Increased doses of melatonin increased leaf dry weight (DLW) at 200 mM salt stress conditions. However, no similar effect was observed in L-tr doses.

External applications had no effect on the surface temperature of the leaves. The difference between these values was statistically the same at 0 and 100 mM salt concentrations. LST increased slightly at 200 mM salt concentration. The results obtained in the leaf length (LL), leaf width (LW), and mean leaf surface area (MLSA) values are similar to LST. Exogenous applications did not have any effect on these results. Only at the highest salt

dose, negative changes were observed. The effects of exogenous melatonin applied to lettuce seedlings under salt stress were investigated by Wie et al. (2018). At the end of the study, it was determined that spraying melatonin on the leaves increased the stomatal conductance, transpiration rate, and net photosynthetic rate of the plants. The act of melatonin on the photosynthetic features involved in nitrate assimilation under the excess nitrate stress in lettuce was detected by Zhou et al. (2019). The findings indicated that supplementation of melatonin decreased the nitrate content both in leaves and roots. Also the fresh and dry weights of seedlings were dramatically increased depending on melatonin.

There are few studies investigating the effects of L-tr on plants. In the study of Khan et al. (2019), the effect of three different amino acids (L-methionine, L-glycine, and L-tryptophan) on different growth parameters of lettuce was investigated. L-methionine significantly increased the growth performance, whereas growth using L-tryptophan and L-glycine decreased.

Even though varied reports about the effect of Mel and L-tr on growth parameters of different plant species were published, this is the first study in which two subtracts are tested together under the salinity conditions for lettuce. The highest stress condition in the study was determined as irrigation with 200 mM NaCl using previous studies. No diseases, pests or physiological disorders were encountered during cultivation except for those caused by salt stress. At the end of the study, the highest dose of melatonin and L-Tryptophan had the highest positive effect on the number

of leaves, salinity necrosis, fresh leaf weight, fresh root weight, and total surface area of lettuce plants under severe stress conditions. The effect of melatonin in all these traits was higher than L-Tryptophan. However, since the number of doses was limited to three, the results could not be tested in higher doses. Therefore, further studies need to determine higher doses. In the dry weight values of the leaves, only the highest dose of melatonin had a positive effect. Results of root dry weight were different than the others. For this feature, the middle dose of melatonin showed the highest effect, but there was a serious decrease in the increasing dose. Some physical dimensions of the leaves (width and length) decreased only depending on the dose of salt stress but were not affected by external applications. Similarly, leaf surface temperature only reacted to salt stress. These results need to be examined further by checking environmental factors. In addition, in order to reach a more general judgment, the dose range of external applications should be expanded and the number of genotypes /varieties used should be increased.

### Acknowledgments

This work was supported by the Research Fund of the Erciyes University. Project Number: FLO-2018- 8572.

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